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MXL-3 and HLH-30 transcriptionally link lipolysis and autophagy to nutrient availability

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Fat is stored or mobilized according to food availability. Malfunction of the mechanisms that ensure this coordination underlie metabolic diseases in humans. In mammals, lysosomal and autophagic function is required for normal fat storage and mobilization in the presence and absence of food. Autophagy is tightly linked to nutrients. However, if and how lysosomal lipolysis is coupled to nutritional status remains to be defined. Here we present MXL-3 and HLH-30 as transcriptional switches coupling lysosomal lipolysis and autophagy to nutrient availability and controlling fat storage and ageing in *Caenorhabditis elegans*. Transcriptional coupling of lysosomal lipolysis and autophagy to nutrients is also observed in mammals. Thus, MXL-3 and HLH-30 orchestrate an adaptive and conserved cellular response to nutritional status and regulate lifespan.

The importance of lipolysis to general metabolism became apparent when it was discovered that fat can only be mobilized in its hydrolysed form[1]. Classically, lipolysis has been associated with cytosolic or ER-associated neutral lipases. However, cytosolic lipids are also catabolized through autophagy-mediated lipolysis, also termed lipophagy[2]. Lysosomal acid lipases[3] (LALs) and Atg15 (ref. 4), an autophagy protein with predicted triglyceride-lipase activity, have been proposed to catabolize lipid-droplet fat stores through lipophagy. Although lysosomal inhibitors induce increased levels of lipid-droplet fats, lysosomal inhibition impairs the function of the endocytic pathway and autophagy, making this approach insufficient to define which lipases actually break down lipids through lipophagy. Similar to general macroautophagy, lipophagy is activated by fasting. However, it is unknown if the assembly of autophagosomes is the only regulated step or if the lipases breaking down lipids in the autophagolysosome would be separately regulated by changes in nutrient availability. If the lipases were independently regulated, then the molecular players that link their activity to nutrient availability would remain to be uncovered. Finally, it has not been established if lipophagy is confined to mammals or if it is an ancient mechanism of energy homeostasis.

Here we present the lysosomal lipases LIPL-1 and LIPL-3 as key enzymes breaking down lipid-droplet fats through lipophagy in *C. elegans*. In addition, we show that the transcription factors MXL-3 and HLH-30 link nutrient availability to lysosomal lipolysis, uncovering two fat regulators. We also show that lysosomal lipolysis and autophagy are similarly linked to nutrient availability in mammalian cells in culture. Finally, we show that lysosomal lipolysis and its regulators MXL-3 and HLH-30 influence *C. elegans* ageing.

RESULTS

lipl-1, *2*, *3*, *4* and *5* respond to fasting

A list of 84 conserved *C. elegans* genes likely to be regulated by nutritional status was generated through comparative analyses of the transcriptional response to starvation of *C. elegans*[5], *Drosophila melanogaster*[6], and mice[7] (Supplementary Tables S1 and S2). We tested these genes for differential expression in 6 h-fasted versus well-fed fertile adult worms. The most upregulated genes of the set belonged to a family of predicted triglyceride lipases. *lipl-1*, *2*, *3*, *4* and *5* were upregulated 44-, 12-, 25-, 7- and 2-fold, respectively (Fig. 1). *lipl-6*, *7* and *8* showed no change in expression (Fig. 1). Transcriptional reporters for *lipl-1*, *2* and *3* are observed in the intestine of fasted, but not well-fed, larvae and adults. The *lipl-4* P::GFP (green fluorescent protein) fusion gene is expressed in the intestine of starvation-induced dauer larvae, and in the pharynx of both well-fed and fasted animals (Supplementary Fig. S1). *lipl-5* P::GFP is expressed in the intestine[8].

LIPL-1 and LIPL-3 are lysosomal lipases controlling lipid-droplet fats

The *lipl* genes encode for uncharacterized triglyceride lipases with extensive sequence similarity to human lysosomal acid lipase (BLAST scores 9e-78 and 3e-75 for *lipl-1* and *lipl-3*, respectively). The primary sequence of the fasting-responsive LIPL proteins contains predicted signals for lysosomal localization (http://golgi.unmc.edu/ptarget/), with confidence scores of 87%, 75%, 62% and 56% for LIPL-1, 3, 2 and 5, respectively. LIPL-4 was not predicted to be lysosomal[9,10]. We characterized the body distribution and subcellular localization of LIPL-1, 2 and 3. Translational fusions of *lipl-1*, *2* and *3* to

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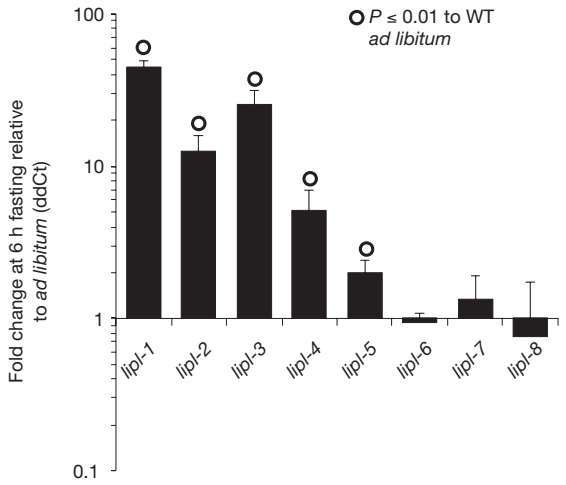

Figure 1 *lipl-1* to *5* are upregulated ~~on~~ fasting. RNA was extracted from young fertile adults fed *ad libitum* or fasted for 6 h. ddCts were calculated normalizing to *ama-1* and the efficiency of the primer sets as previously described[28]. Means+s.e.m. are depicted. *lipl-1* to *lipl-5* data are derived from six independent experiments. *lipl-6* to *lipl-8*, $n=3$ independent experiments. Significant *t*-test derived *P*-values are indicated. WT, wild type.

either GFP or TagRFP (red fluorescent protein) reveal expression of these proteins in the lumen of the gut and/or vesicles within the intestine (Supplementary Fig. S2a). Confocal microscopy shows that LIPL-1 and 3 co-localize with the lysosomal marker PGP-2 (ref. 11; Fig. 2a). Furthermore, fractionation of whole lysates of worms expressing LIPL-3::TagRFP shows that both the acid lipase activity and LIPL-3::TagRFP co-fractionate with the canonical lysosomal enzyme acid phosphatase (Supplementary Fig. S2b), confirming that LIPL-1 and 3 localize to the lysosomal-related organelle of *C. elegans*. Moreover, *lipl-1(tm1954) lipl-3(tm4498)* double mutant worm lysates have reduced acid lipase activity (pH 4.5; Fig. 2b), but normal neutral lipase activity (Supplementary Fig. S2c), suggesting that LIPL-1 and 3 are only active in the acidic conditions that characterize the lysosomal lumen.

Lysosomal lipases have been proposed to break down lipid-droplet fats[3]. We investigated the impact of inactivating *lipl-1* and *lipl-3* on the accumulation of cytoplasmic fats. *lipl-1* and *lipl-3* double mutant larvae show threefold greater fat stores than wild-type larvae (Fig. 2c). A significant increase in fat signal in adult animals is also observed (Supplementary Fig. S3a). Furthermore, dual inactivation of *lipl-1* and *lipl-3* impaired fat utilization ~~on~~ fasting (Fig. 2d). Transmission electron microscopy confirms that *lipl-1 lipl-3* double mutant animals contain while feeding, and retain during fasting, more and larger lipid droplets than wild-type animals (Fig. 2e and Supplementary Fig. S3b), further suggesting that *lipl-1* and *3* break down lipids contained in lipid droplets. However, the LIPL lipases do not co-localize with lipid droplets (Fig. 2f).

In mammals, in basal and fasting conditions, autophagy delivers lipid-droplet lipids to the lysosome: a process termed lipophagy[2]. In *C. elegans*, dual inactivation of the essential autophagy genes *lgg-1* and *lgg-2* (LC3 homologues) leads to increased fat accumulation ~~in worms~~ (Fig. 2g), suggesting that lipophagy is conserved across metazoans. Lysosomal[3] and autophagic[4] lipases were proposed to break down lipid-droplet fats through lipophagy. Inhibition of autophagy by RNA interference (RNAi) of *lgg-1* and *lgg-2* in the *lipl-1 lipl-3* double mutant animals does not lead to further increases in lipid-droplet fat stores (Fig. 2g), confirming that these lysosomal lipases and autophagy are part of the same fat regulatory pathway and suggesting that LIPL-1 and LIPL-3 are the enzymes breaking down lipids through lipophagy in *C. elegans*. By contrast, no changes in the levels of expression of lipid synthesis or β-oxidation genes were observed in *lipl-1 lipl-3* double mutant animals (Supplementary Fig. S3c).

MXL-3 represses the *lipl* genes when food is available

To identify the transcription factors that link lysosomal lipase messenger RNA levels to nutrient availability, a GFP transcriptional fusion to the *lipl-1* promoter (*lipl-1*-P::GFP) was used to screen an RNAi sublibrary containing 403 predicted *C. elegans* transcriptional regulators and 193 nuclear hormone receptors (Supplementary Table S3). Inactivation of the basic-helix–loop–helix transcription factor *mxl-3* (Max-like 3) enabled *lipl-1*-P::GFP expression under well-fed conditions. Gene expression analyses of two *mxl-3* null mutants, *mxl-3(ok1947)* and *mxl-3(tm2580)*, confirmed that MXL-3 represses *lipl-1*, *2*, *3* and *lipl-5*, but not *lipl-4*, in well-fed animals (Fig. 3a). *mxl-3* mRNA levels are also responsive to fasting, dropping by up to 15-fold after 5 h of fasting (Fig. 3b), suggesting that *mxl-3* inactivation is part of a physiological response to fasting. Levels of *mxl-3* transcripts return to 30% of those observed in well-fed animals after 12 h of fasting. Similarly, the *lipl* genes return to near basal levels after 18 h of fasting, suggesting that MXL-3 orchestrates a transient response to fasting. A ~~functional~~ MXL-3::GFP fusion protein, which rescues the *mxl-3(ok1947)* transcriptional phenotype, localizes to the nuclei of intestinal cells and ~~chemo~~sensory neurons (Supplementary Fig. S4a–c). Tissue-specific inactivation of *mxl-3* in the gut, rather than neurons, triggered a fasting-like transcriptional response (Supplementary Fig. S4d). MXL-3::GFP expressed from a low-copy array localized to the nucleus of well-fed animals, but the signal disappeared from the nuclei after 2 h of fasting and remained undetectable during 6 h of fasting (Fig. 3c and Supplementary Fig. S4e). Twenty-two and 68% of intestinal cells showed nuclear-localized MXL-3::GFP after 12 and 18 h of fasting, respectively (Fig. 3c and Supplementary Fig. S4e). The transient regulation of MXL-3 suggests that the transcriptional changes coupled to inactivation of MXL-3 are part of an acute, rather than prolonged, response to nutritional deprivation. The observation that the *lipl* genes and *mxl-3* have opposite patterns of expression and that they are both expressed in the intestinal cells supports the hypothesis that MXL-3 directly represses the *lipl* genes. Furthermore, MXL-3 binds *in vitro* to CACGTG (ref. 12), and this target motif is present within 500 base pairs (bp) of the transcriptional start sites of *lipl-1*, *2*, *3* and *5*, but not in the promoter region of the fasting-responsive gene *lipl-4*, whose transcription is *mxl-3* independent. Also, MXL-3 binds and drives expression from the promoter regions of *lipl-1*, *2* and *3*, but not *lipl-4* in yeast one-hybrid experiments (Supplementary Fig. S5a). Chromatin immunoprecipitation (ChIP) of MXL-3::GFP shows that MXL-3 binds *in vivo* to the promoters of *lipl-1* and *3* in well-fed animals and that MXL-3 occupancy of these promoters drops by between 5- and ~~50~~-fold in fasting worms (Fig. 3d and SI_ChromatinIP). In summary, MXL-3 acts as a nutritionally regulated transcriptional repressor of an acute response to fasting that includes induction of the lysosomal lipase genes *lipl-1* and *3*.

Acute inactivation of *mxl-3* through RNAi leads to a depletion of fat stores that is comparable to the levels of fat consumption induced

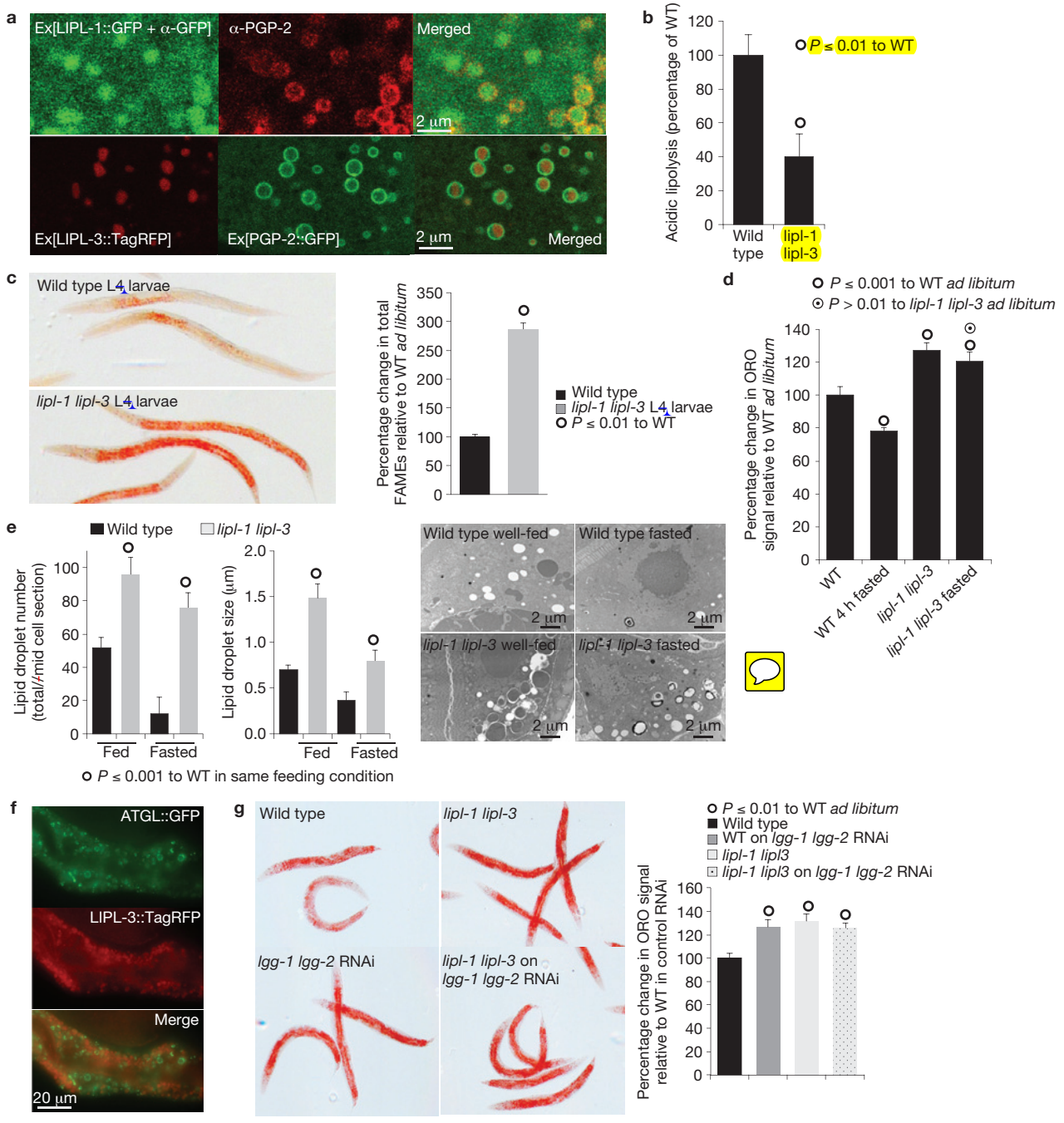

Figure 2 LIPL-mediated lysosomal lipolysis controls lipid-droplet fat stores. (**a**) Representative confocal images of LIPL-1::GFP and LIPL-3::TagRFP localization relative to the lysosomal marker PGP-2 show that LIPL-1 and LIPL-3 are localized to the lysosomal-related organelle. (**b**) Acid lipase activity, measured in 1-day adult whole lysates, shows that *lipl-1 lipl-3* double mutant animals have reduced acidic lipolytic capacity. Mean ± s.d. are presented relative to wild type (WT); significant differences are indicated; $n = 4$ independent experiments. (**c**) Oil Red O (ORO) staining and fatty acid methyl ester (FAME) analyses of wild-type and *lipl-1(tm1954) lipl-3(tm4498)* double mutant L4 larvae show that lysosomal lipases regulate cytosolic fat stores (see Fig. 2d for adult measurements). Means ± s.e.m. are presented relative to wild type; significant differences are indicated; $n = 3$ independent experiments. (**d**) ORO quantification (mean percentage ± s.e.m. relative to wild type ~~on vector control~~) of wild-type and *lipl-1(tm1954) lipl-3(tm4498)* double mutant young adults fasted for 4 h shows that LIPL-1 and LIPL-3 contribute to fat mobilization ~~on~~ fasting. Wild-type fasted worms show 20% less ORO signal than animals fed *ad libitum* ($P \leq 0.001$), whereas *lipl-1(tm1954) lipl-3(tm4498)* double mutant worms show 7% reduction in ORO signal (a difference that is not significant to well-fed fat levels at $P \leq 0.01$ but it is significant at $P \leq 0.05$). $N = 3$ independent experiments. (**e**) Representative transmission electron microscopy images of well-fed and 6 h-fasted wild-type and *lipl-1 lipl-3* mutant animals (11,500×) show that the *lipl* mutants have more lipid-droplet stores in *ad libitum* fed and fasted conditions. Quantification of vesicle number and size is depicted ~~below the images~~ as mean ± s.e.m., $n = 4$ independent experiments. (**f**) Representative ~~immunostaining against TagRFP and GFP~~ of well-fed transgenic animals expressing LIPL-3::TagRFP and ATGL::GFP shows that LIPL-3 does not localize to lipid droplets. (**g**) ORO staining and quantification (mean percentage ± s.e.m. relative to wild type on vector control) of wild-type and *lipl-1(tm1954) lipl-3(tm4498)* double mutant young adults treated post-developmentally (from L4) with RNAi against the essential autophagy genes *lgg-1* and *lgg-2* or vector control show that *lipl-1 lipl-3* and the autophagy genes *lgg-1 lgg-2* are in the same fat regulatory pathway. $N = 4$ independent experiments.

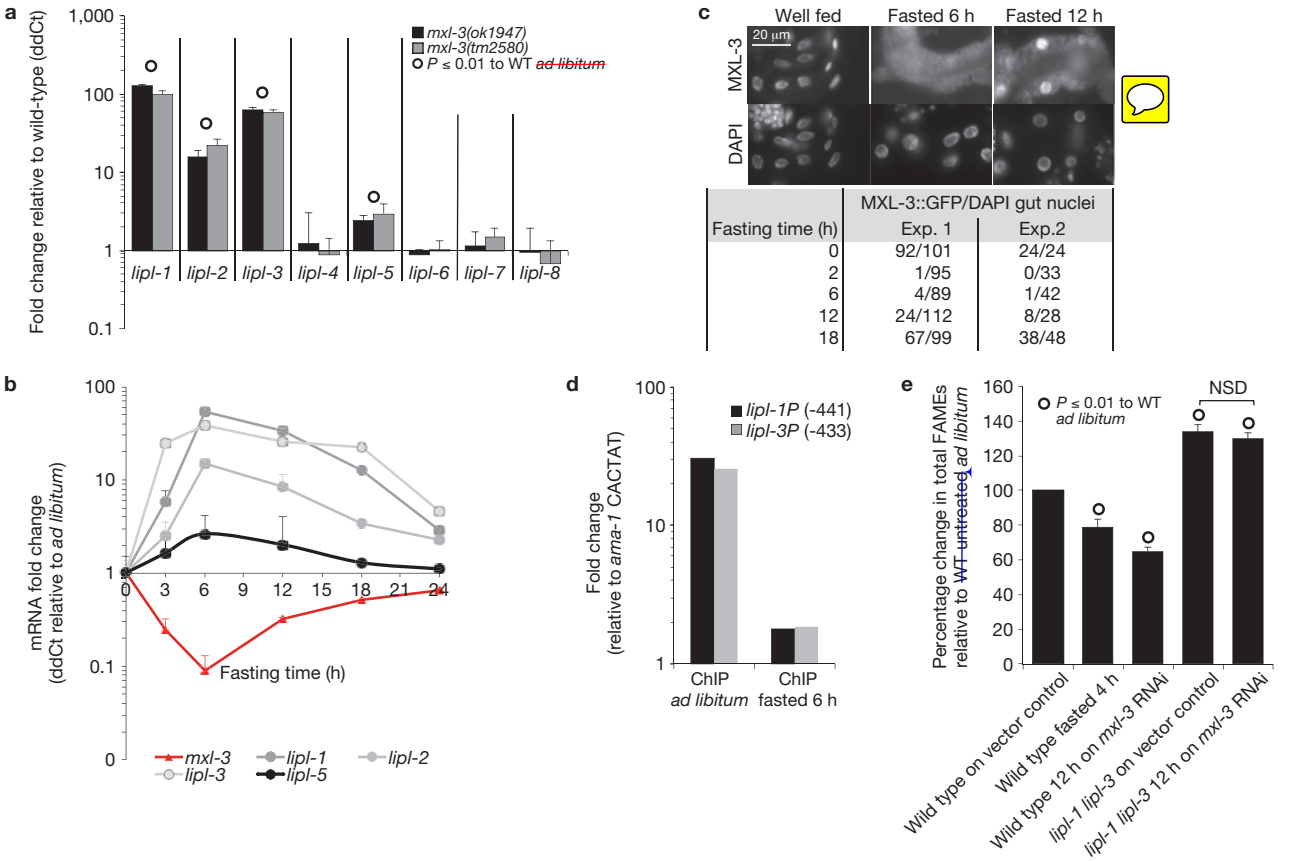

Figure 3 MXL-3 represses lysosomal lipolysis in *ad libitum*-fed conditions. (**a**) Expression of *lipl* genes in well-fed *mxl-3(ok1947)* and *mxl-3(tm2580)* young adults normalized to same age well-fed wild-type (WT) worms shows that *mxl-3* loss of function is sufficient to induce *lipl-1* to *3* and *lipl-5*. Mean ddCts+s.e.m. are depicted. $N=6$ independent experiments for *lipl-1* to *lipl-5*, and $n=4$ independent experiments for *lipl-6* to *lipl-8*. (**b**) L4 larvae were fasted and RNA was extracted at the indicated times. Mean ddCts+s.e.m. show that the response *mxl-3* orchestrates is transient. Time 0, $n=6$ independent experiments; time 3–12 h, $n=4$ independent experiments; time 18–24 h, $n=2$ independent experiments. (**c**) Immunostainings of well-fed or 6 or 12 h fasted MXL-3::GFP young adults are presented. Quantification of GFP-positive nuclei relative to total intestinal nuclei (4,6-diamidino-2-phenylindole, DAPI) of two independent experiments shows that MXL-3 transiently delocalizes from the intestinal nuclei during fasting. (**d**) ChIP–quantitative PCR (ChIP-qPCR) analysis of well-fed and 6 h fasted mixed-stage worms expressing MXL-3::GFP presented as Ct in αGFP immunoprecipitated DNA normalized to input DNA and relative to a mock promoter region (CACTAT site 88 of *ama-1* gene) shows that MXL-3 vacates the *lipl* promoters during early fasting. Three sets of primers surrounding CACGTG target sites found ~~that~~ up to 500 bp of the ATG of the *lipl-1* and *lipl-3* genes were used. A representative experiment is presented; see raw data of two independent experiments in Supplementary Information. (**e**) Wild-type or *lipl-1(tm1954) lipl-3(tm4498)* double mutant animals grown on control RNAi plates were transferred as L4 larvae to control or *mxl-3* RNAi plates, incubated for 12 h at 20 °C, and processed for total FAMEs; as a reference, an aliquot of wild-type young adults on control RNAi bacteria was fasted for 6 h. FAME quantification (mean percentage of fed wild type ±s.e.m.) shows that acute *mxl-3* inactivation, as fasting, reduces fat stores in a *lipl*-dependent manner. $N=3$ independent experiments. NSD, *P*-value > 0.05.

by fasting (Fig. 3e). Although it is possible that *mxl-3* inactivation also affects β-oxidation, *mxl-3* mutant animals show wild-type levels of oxygen consumption (Supplementary Fig. S5c), ~~suggesting that β-oxidation is not a major metabolic change associated with *mxl-3* loss of function~~. By contrast, the fast fat depletion induced by acute *mxl-3* inactivation depends on the activity of the lipases LIPL-1 and 3 (Fig. 3e). Taken together, the data support the model that MXL-3 is a key molecular switch, whose inactivation turns on a fat-mobilizing program in response to fasting, and that the LIPL lipases are major players executing this metabolic program.

We then asked if certain well-characterized nutrient sensors of *C. elegans* regulated the expression of *mxl-3*. Inactivation of CeTOR (target of rapamycin) through RNAi or transforming growth factor (TGF-) β (*daf-7(e1372)*) did not affect *mxl-3* expression (Supplementary Fig. S5d,e). On the other hand, impaired insulin signalling (*daf-2(e1368)*) leads to modest (50%) repression of *mxl-3* transcription and six- and twofold induction of *lipl-1* and *lipl-3*, respectively (Supplementary Fig. S5f), suggesting that reduced insulin signalling is only part of the regulatory cascade that links MXL-3 function to nutrients.

HLH-30 induces the *lipl* genes ~~on~~ fasting

A candidate gene approach was used to identify transcription factors that activate the *lipl* genes in response to fasting. MXL-3 has the same binding site and shares target genes with HLH-30 (ref. 12), thus we investigated the role of *hlh-30* in the activation of lysosomal lipolysis in fasting worms. In contrast to *mxl-3*, *hlh-30* transcription is induced in fasting worms (Fig. 4a), and repressed ~~on~~ refeeding (Supplementary Fig. S6a). Inactivation of *hlh-30* by a chromosomal mutation, allele *tm1978*, delays the activation of an *hlh-30* transcriptional fusion to GFP (Supplementary Fig. S6b), suggesting that HLH-30 positively regulates its own transcription. Inactivation of CeTOR induces the expression

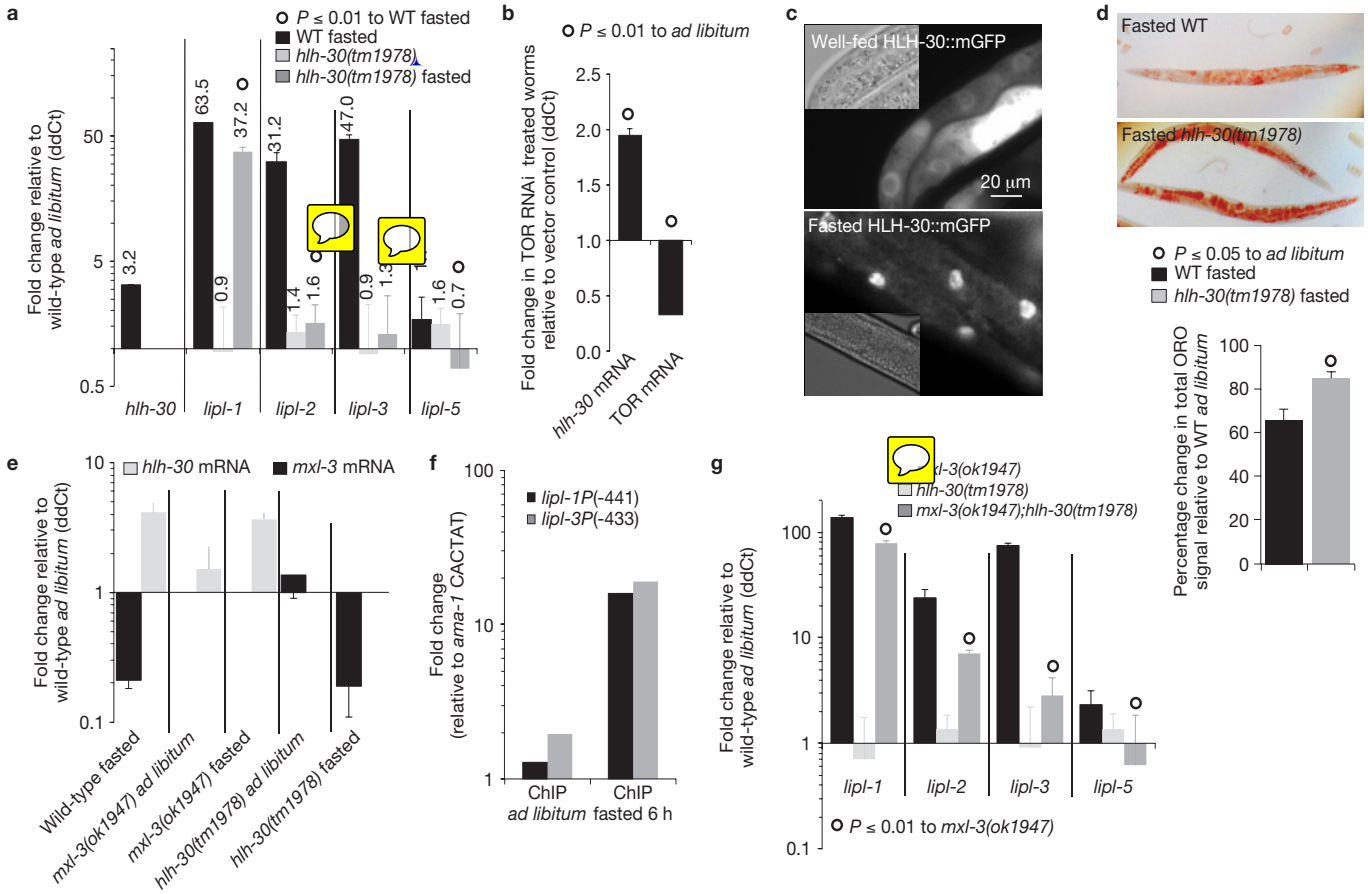

Figure 4 HLH-30 induces lysosomal lipolysis ~~on~~ fasting. (**a**) Transcriptional levels of *hlh-30* and the *mxl-3*-dependent *lipl* genes measured in 5 h fasted wild-type (WT) or *hlh-30(tm1978)* young-adult worms depicted as mean ddCt ± s.e.m. relative to wild-type *ad libitum*-fed worms show that *hlh-30* is induced ~~on~~ fasting and HLH-30 induces lysosomal lipolysis. $N = 4$ independent experiments for *lipl* data, $n = 2$ independent experiments for *hlh-30* mRNA. HLH-30 deficiency fully abrogates *lipl-2*, *3* and *5* ($P < 0.0001$), and impairs *lipl-1* ($P < 0.01$) transcriptional activation ~~on~~ fasting. (**b**) Animals treated from late L3 stage with RNAi against TOR or vector control were harvested as young adults. ~~ddCts calculated and presented as in Fig. 1~~ show that inhibition of TOR is sufficient to induce *hlh-30* transcription. Means ± s.e.m. of three independent experiments ~~are depicted~~. (**c**) L3 animals carrying the rescuing construct *hlh-30* P::HLH-30::~~e~~GFP::*hlh-30* (eGFP, enhanced GFP) 3′UTR (untranslated region), well fed or fasted for 8 h show that HLH-30 is enriched in intestinal nuclei of fasted worms (exposure time: well fed, 500 ms; fasted, 100 ms). (**d**) ORO staining of wild-type or *hlh-30(tm1978)* young adults fasted for 8 h reveals that *hlh-30* is required for optimal lipid mobilization on fasting. Mean percentages ± s.e.m. are shown relative to well-fed worms treated in parallel, $n = 3$ independent experiments. (**e**) *mxl-3* and *hlh-30* transcriptional levels in wild-type, *hlh-30(tm1978)* and *mxl-3(ok1947)* mutants in the basal and fasted states show that *mxl-3* transcription is *hlh-30* independent, and *hlh-30* transcription is *mxl-3* independent. Expression is presented relative to wild-type fed animals as mean ddCt ± s.e.m. No significant differences relative to wild type in the same feeding state were observed, $n = 3$ independent experiments. (**f**) Representative ChIP-qPCR analysis of well-fed and 6 h fasted mixed stage worms expressing HLH-30::GFP presented as in Fig. 3d shows that HLH-30 occupies the *lipl* promoters during early fasting. See raw data of two independent experiments in Supplementary Information. (**g**) Transcriptional levels of the *mxl-3*-dependent *lipl* genes in wild-type, *mxl-3(ok1947)*, *hlh-30(tm1978)* or *mxl-3(ok1947);hlh-30(tm1978)* double mutant young-adult worms show that *hlh-30* suppresses the constitutive-induction-of-the-*lipl*-genes phenotype of *mxl-3* mutant animals (mean ddCt ± s.e.m. relative to wild-type *ad libitum*-fed worms). $N = 3$ independent experiments.

of *hlh-30* (Fig. 4b), revealing that mTOR controls *hlh-30/CeTfeb* at the transcriptional level. On fasting, HLH-30 is enriched in the nuclei (Fig. 4c). In turn, *hlh-30* induces *lipl-2*, *3* and *5* expression during fasting, and contributes to the induction of *lipl-1* (Fig. 4a), suggesting that *hlh-30* would control lipid mobilization ~~on~~ fasting. As expected, *hlh-30* mutants show a reduced capacity to mobilize cytosolic lipids after food withdrawal (Fig. 4d). Mutational inactivation of *hlh-30* does not affect either the basal transcription or fasting-induced repression of *mxl-3*. Conversely, mutation of *mxl-3* does not affect either the basal transcription or fasting-triggered induction of *hlh-30* (Fig. 4e). These findings show that *mxl-3* and *hlh-30* transcripts are independently regulated. Nevertheless, as analysis by ChIP shows that HLH-30 is rarely associated with the promoters of the lipase genes in well-fed animals (Fig. 4f and SI_ChromatinIP), even when the *hlh-30* gene is expressed from a high copy number array at higher levels than those observed in fasting animals (Supplementary Fig. S6c), the coordinated relay from repression by MXL-3 to activation by HLH-30 seems critical for the proper expression of the lipase genes in response to nutrient availability. Consistent with this hypothesis, *lipl-1* and *3* expression is only slightly increased in well-fed animals overproducing HLH-30 (Supplementary Fig. S6c). In contrast, during fasting, when MXL-3 is not bound to the lipase promoters, HLH-30 occupies the promoters of the lipase genes and induces expression of these targets (Fig. 4f and Supplementary Fig. S6c). Moreover, loss of *hlh-30* function suppresses

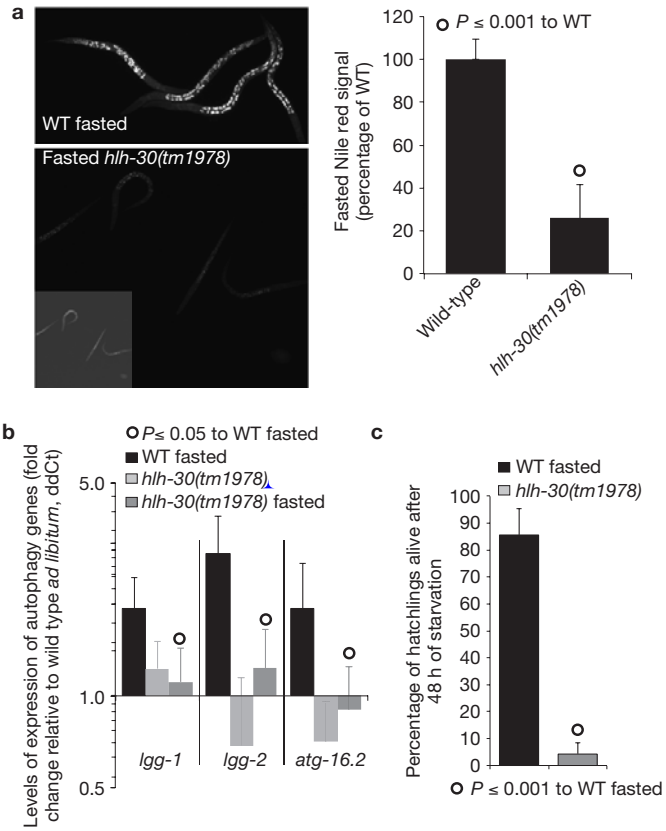

Figure 5 HLH-30 activates vital cellular responses to starvation. (**a**) LRO live Nile red-stained images of *hlh-30(tm1978)* and wild-type (WT) L3 larvae starved for 60 h show that *hlh-30* is required for the expansion of the lysosomal compartment ~~on~~ fasting. Quantification as mean ± s.e.m. relative to fasted wild-type worms is also presented; $n = 3$ independent experiments. (**b**) Transcriptional levels of the autophagy genes *lgg-1*, *lgg-2* and *atg-16.2* in fasted wild-type and *hlh-30(tm1978)* young-adult worms show that transcriptional activation of autophagy ~~on~~ fasting is *hlh-30* dependent (mean ddCt ± s.e.m. relative to wild-type *ad libitum*-fed worms). $N = 3$ independent experiments. (**c**) The mean percentage (±s.d.) of L1 larvae alive after 48 h fasting in minimum media shows that *hlh-30* mutant animals are sensitive to starvation. $N = 3$ independent experiments. For L4 survival see Supplementary Fig. S6g.

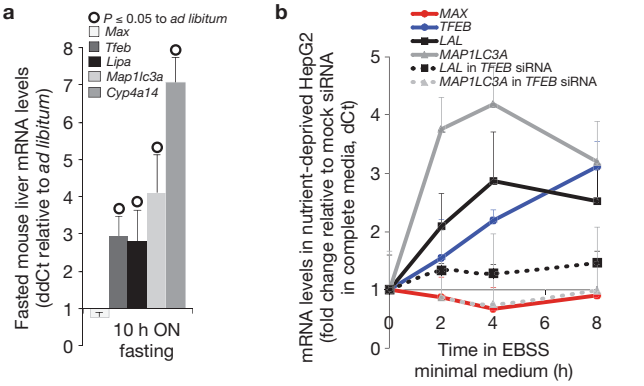

Figure 6 Lysosomal lipolysis and autophagy are transcriptionally linked to nutrients in mammals. (**a**) Expression analyses of the liver of C57BL/6J 9-week females fasted overnight (ON) for 10 h relative to siblings feeding *ad libitum* show that *LipA* (mouse lysosomal acid lipase), *Map1lc3a* (mammalian *lgg-1*/*lgg-2*) and *Tfeb* (mammalian *hlh-30*) but not *Max* (mammalian *mxl-3*) are transcriptionally linked to nutrients in the mouse liver (median ± s.e.m. fold change; ddCt). Levels of expression were normalized to *ActB* and *Cog2* as internal controls; all but *Max* differences are significant ($P < 0.05$), $n = 3$ independent experiments. *Cyp4a14* is a positive control. (**b**) Expression analyses of control or h*TFEB* siRNA treated HepG2 cells incubated in ~~EEBS~~ minimal medium for 2, 4 or 8 h, compared with the expression of control or h*TFEB* siRNA treated hepatocytes in complete media, show that *LAL* (human lysosomal acid lipase), *MAP1LC3A* and *TFEB* but not *MAX* are transcriptionally linked to nutrients in human hepatocytes, and *LAL* and *MAP1LC3A* induction under nutrient deprivation are *TFEB* dependent (median dCt ± s.e.m.). Levels of expression were normalized to *ACTB* as internal control; all but *MAX* differences are significant ($P < 0.05$), $n = 3$ independent experiments. *IGFBP* is a positive control.

the constitutive induction of the lipase genes observed in *mxl-3* mutants (Fig. 4f). Taken together, these findings show that *mxl-3* and *hlh-30* antagonistically regulate lysosomal lipase gene expression in response to nutrient availability. Finally, adaptive induction of the *lipl* genes in fasting animals is independent of the metabolic regulators NHR-49 and SIR-2.1 (Supplementary Fig. S6d,e).

During fasting the *C. elegans* lysosomal compartment expands[13]. Inactivation of *hlh-30* causes a reduction of the live Nile red-labelled lysosomal compartment in well-fed (ref. 14 and Supplementary Fig. S6f) and fasting conditions (Fig. 5a), suggesting that, as does its mammalian counterpart[15,16], *hlh-30* links lysosomal biogenesis to nutrient availability in *C. elegans*.

Autophagy is also activated ~~on~~ fasting[17], and we had observed that autophagy genes are transcriptionally upregulated in fasted animals, so we tested the role of *hlh-30* in the induction of autophagy genes. Fasting *hlh-30* mutants fail to activate the transcription of essential autophagy genes (Fig. 5b and Supplementary Table S4). The results support the model that HLH-30 coordinates the activation of lysosomal lipolysis and autophagy to meet the nutritional needs of the cells. Notably, *hlh-30* mutant animals die prematurely in starvation conditions (Fig. 5c and Supplementary Fig. S6g), demonstrating that *hlh-30* is required to mount an essential metabolic response to starvation.

Essential metabolic responses to food deprivation are expected to be under strong selective pressure and consequently conserved. Supporting the view that lysosomal lipolysis is also regulated by nutrients in mammals, increased acid lipase activity was observed in liver lysates from fasted, compared with well-fed, C57L/B6 mice (Supplementary Fig. S7a). Furthermore, TFEB, LAL and the essential autophagy gene product LC3 were induced both in the liver of fasted C57L/B6 mice and in serum-deprived HepG2 cells (Fig. 6a,b), suggesting that lysosomal lipolysis and autophagy may be controlled by similar transcriptional mechanisms across metazoans. Confirming this hypothesis, siRNA inactivation of TFEB impairs the transcriptional activation of LAL and LC3 in nutrient-deprived HepG2 cells (Fig. 6b). In contrast, expression of the closest mammalian homologue of *mxl-3*, *MAX*, was not repressed in either fasted mice or serum-deprived hepatocytes (Fig. 6,b). Nonetheless, expression of mouse *MAX* in *C. elegans* rescued the constitutive induction of lipase genes in *mxl-3* mutants (Supplementary Fig. S7b), suggesting that MAX, or a close member of the MAX family, could regulate energy homeostasis in higher organisms.

MXL-3 and HLH-30 regulate *C. elegans* ageing

mxl-3 mutant animals show a constitutive fasting-like transcriptional profile, specifically, the constitutive activation of the lysosomal lipases. Similar to animals subjected to low nutrient intake, *mxl-3* mutant

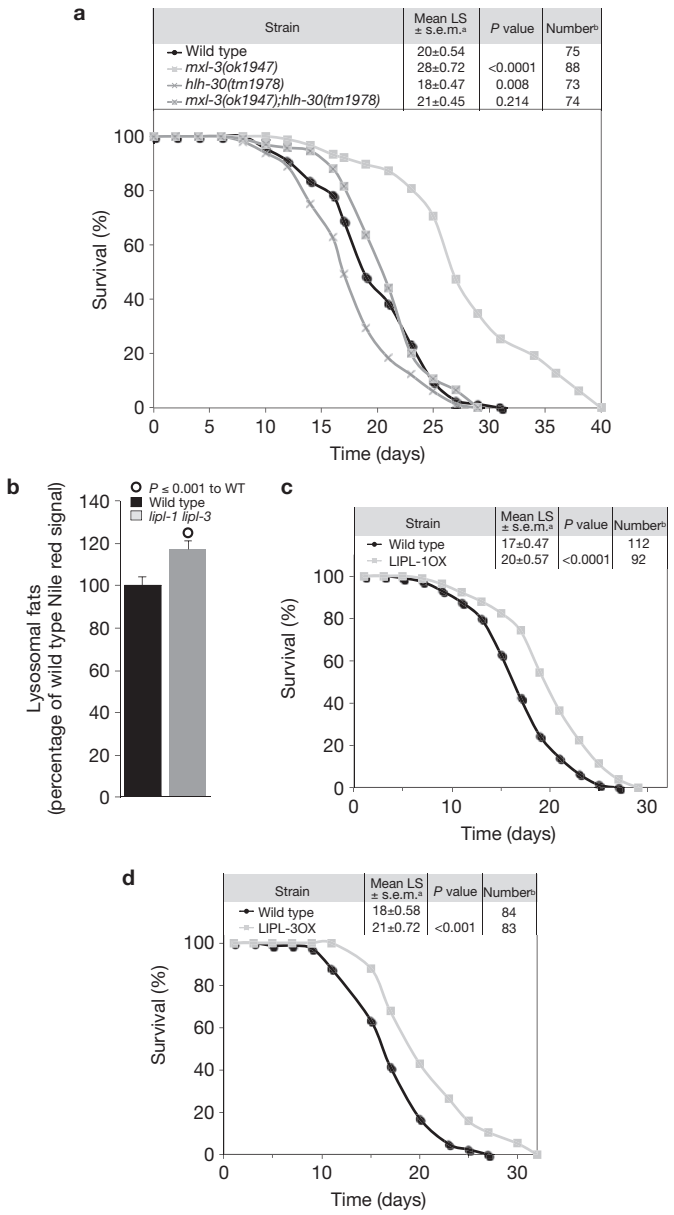

Figure 7 Lysosomal lipolysis delays *C. elegans* ageing. (**a**) Wild-type (WT), *mxl-3(ok1947)*, *hlh-30(tm1978)* or *mxl-3(ok1947);hlh-30(tm1978)* double mutant were used in longevity epistasis analyses, revealing that *hlh-30* suppresses the *mxl-3* extended lifespan phenotype. Animals were incubated at 20 °C and transferred every other day until cessation of reproduction. The cumulative survival curve is depicted. Kaplan–Meier statistics, calculated using SSPS 17, are also shown. (**b**) Live Nile red quantification of wild-type and *lipl-1 lipl-3* double mutant worms shows that *lipl-1* and *lipl-3* contribute to the clearance of lipids from the endocytic pathway. Mean ± s.e.m. signal intensity is shown as percentage relative to wild type; significant differences are indicated; $n = 4$ independent experiments. (**c**,**d**) Animals overexpressing LIPL-1 or LIPL-3 were incubated at 25 °C and transferred every other day until cessation of reproduction. **a** Survival presented as mean lifespan (LS) ± s.e.m. **b** Number of uncensored animals (animals that crawled off the plate, bagged, exploded or became contaminated were censored). Data presented as in Fig. 7a show that activated lysosomal lipolysis extends *C. elegans* lifespan.

worms are long lived in abundant food conditions (Fig. 7a). The *mxl-3* gene is expressed in the intestine and AWC-sensory neurons (Supplementary Fig. S4). *mxl-3* mutants undergo chemotaxis normally towards the AWC-specific odorant isoamyl alcohol (Supplementary Fig. S8a), suggesting that this mutant has an intact chemosensory system and that its extended lifespan phenotype is not due to impaired sensory abilities. Furthermore, intestinal-specific expression of *mxl-3* rescues the *mxl-3* extended lifespan phenotype (Supplementary Table S5). The *mxl-3* mutant long-lived phenotype is additive to the lifespan extension derived from deficient germline stem cells (*glp-1(e2141)*), mitochondrial dysfunction (*clk-1* RNAi) and a hypomorphic mutation in the insulin receptor (*daf-2(e1368)*), and it was not suppressed by inactivation of the transcription factors *skn-1(zu67)* or *daf-16(mu86)*, which are required for lifespan extension due to insulin deficiency (Supplementary Table S6). Although *skn-1* and *daf-16* inactivation partially suppressed the extended lifespan phenotype of the *mxl-3* mutant animals, they affect wild-type longevity to the same extent that they affect the lifespan of the *mxl-3* mutant (20–40%), suggesting they are not required for the *mxl-3* extended lifespan phenotype. The *mxl-3* ageing phenotype was also additive to the *eat-2* model of caloric restriction (*eat-2(ad465)*), and it was not suppressed by inactivation of the transcription factor *pha-4* (RNAi), or *rheb-1*, a GTPase required for lifespan extension by intermittent fasting (Supplementary Table S7). Finally, the *mxl-3* extended lifespan phenotype was additive to starvation-induced lifespan extension (Supplementary Table S7), suggesting that *mxl-3* does not represent a genetic model of classical caloric-restriction or starvation-induced longevity.

Conversely, *hlh-30* mutants were modestly short lived (Fig. 7a) and loss of *hlh-30* function suppressed the longevity phenotype of *mxl-3* mutants (Fig. 7a). Targets of *hlh-30* affect both lysosomal function and autophagy, two processes implicated in ageing[18–20]. The *mxl-3* mutants did not show increased autophagy (Supplementary Fig. S8b). Consequently, it is unlikely that *hlh-30* suppresses the *mxl-3* extended lifespan phenotype by impairing autophagic function. In support of this idea, inactivation of the autophagy genes *bec-1* (beclin1 orthologue) or *atg-16.2*, or the transcriptional regulator of autophagy *pha-4*, did not suppress the *mxl-3* longevity phenotype (Supplementary Table S8). The *mxl-3* mutant does not show activated autophagy, but exhibits activated lysosomal lipolysis, and inactivation of *hlh-30* suppressed the induction of lysosomal lipases in *mxl-3* mutants. These findings support the model that increased lysosomal lipolysis extends *C. elegans* lifespan. Efficient clearance of extra- and intracellular components delivered to the lysosome or passing through the endocytic pathway is required for normal ageing[18]. In addition to controlling cytosolic fat stores through lipophagy, the lysosomal lipases LIPL-1 and 3 promote fat mobilization through the endocytic pathway, as made evident by the increased vital Nile red staining observed in *lipl-1 lipl-3* double mutants (Fig. 7b). In agreement with the hypothesis that efficient clearance of lipid moieties from the endosomal/lysosomal compartment slows ageing, overexpression of the *lipl* genes modestly increased *C. elegans* lifespan (Fig. 7c,d). The longevity data therefore suggest that increased lysosomal lipolysis enhances somatic endurance, possibly by improving cellular clearance, re-routing energy reserves for somatic maintenance, or some combination thereof.

DISCUSSION

Our data support the hypothesis that the *C. elegans* homologues of human LAL, the LIPL lipases, mediate the mobilization of cytosolic fats through lipophagy, and that active regulation of lipophagy is

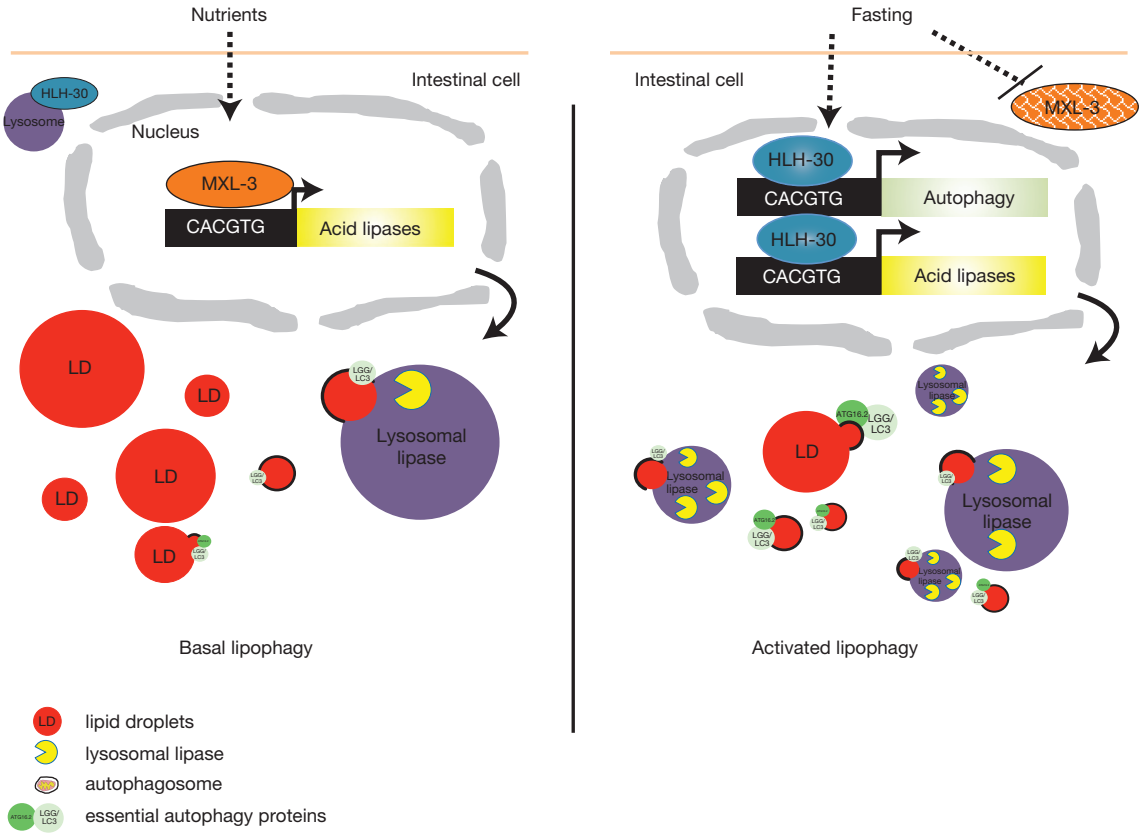

Figure 8 MXL-3 and HLH-30 model of action. In the presence of nutrients, MXL-3 represses the expression of the lysosomal lipase genes, which contribute to breaking down fats through lipophagy. ~~On~~ fasting, MXL-3 is transcriptionally repressed and delocalizes from the nuclei, releasing the repression of the lysosomal lipases. Concomitantly, HLH-30 translocates from the cytoplasm to the nucleus and *hlh-30* expression is induced, in part by an autoregulatory mechanism. In the nucleus HLH-30 enables the activation of lipophagy through the induction of lysosomal lipase and autophagy genes. The activation of this transcriptional program enables utilization of internal reserves of energy, and promotes survival of starvation.

an ancient mechanism of energy homeostasis selected to adapt to fluctuations in food availability. Furthermore, we find that lysosomal lipolysis is tightly linked to nutrient availability through two metabolic regulators, MXL-3 and HLH-30 (Fig. 8). When food is available, MXL-3 represses the transcription of the lysosomal lipase genes. ~~On~~ fasting, *mxl-3* mRNA levels drop and the protein quickly disappears from the intestinal nuclei. Concomitantly, HLH-30 is translocated to the nucleus and the *hlh-30* gene is transcriptionally upregulated, at least in part through an autoregulatory mechanism. This reinforces the activation of a fasting transcriptional program that leads to the transient induction of lysosomal lipolysis and autophagy, which, in conjunction with the canonical neutral lipases[21,22], are required to use internal reserves of energy and survive starvation. Lipid consumption is most important and active in the initial phase of the metabolic response to food deprivation of *C. elegans* males[23]. Here we show that fertile hermaphrodite *C. elegans* transiently activate lysosomal lipolysis, providing a contributing mechanism to the rapid consumption of fat stores in adult worms.

The transcriptional regulation mediated by MXL-3 and HLH-30 functions independently of the metabolic regulators NHR-49 (ref. 24) and SIR-2.1 (ref. 25). Also, although TOR regulates mammalian TFEB protein function[26,27] and *C. elegans hlh-30/CeTfeb* transcription, the inactivation of CeTOR is insufficient to repress *mxl-3*. We show evidence suggesting that the repression of *mxl-3* is required for the activation of its target genes, supporting the hypothesis that in *C. elegans* CeTOR inhibition is insufficient to fully activate this metabolic programme. CeTOR-independent insulin signalling seems to control *mxl-3* expression. However, inhibition of insulin signalling is also insufficient to fully activate lysosomal lipolysis, supporting a model in which multiple nutrient-sensing pathways act upstream of lysosomal lipolysis.

The MXL-3-HLH-30 transcriptional circuit is required not only for proper nutrient mobilization in well-fed and fasted conditions but also for normal ageing. MXL-3 promotes ageing and, at least in the context of the *mxl-3* mutant model, HLH-30 prevents ageing, suggesting that a transcriptional program selected to adapt to fluctuations in food availability regulates ageing when nutrients are available. We show that the *mxl-3* longevity phenotype is additive to caloric restriction and starvation. Although at first surprising, this observation is in agreement with the fact that the response that *mxl-3* orchestrates is transient. Fasting leads to the repression of *mxl-3* and the activation of lysosomal lipolysis, but this response lasts only hours. In contrast, the mean lifespan of post-reproductively starved animals is 29 days. Consequently, it is unlikely that the transient activation of lipophagy would be a major contributor to starvation-induced lifespan extension. Alternatively, we propose that active clearance of lipid moieties from the endosomal/lysosomal compartment and/or signalling molecules derived from lipid breakdown slow ageing in *C. elegans*.

In summary, we show that lysosomal lipolysis is an ancient mechanism of energy homeostasis and we present MXL-3 and HLH-30 as metabolic regulators that orchestrate a conserved and adaptive response to food deprivation, in addition to regulating ageing in conditions of food abundance. Finally, the conservation of the presented response suggests that malfunction of the mechanisms that link nutrients to lysosomal function may underlie metabolic disorders of as yet unknown aetiology and age-related disorders in higher organisms. □

METHODS

Methods and any associated references are available in the online version of the paper.

Note: Supplementary Information is available in the online version of the paper

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AUTHOR CONTRIBUTIONS

E.J.O'R. designed the overall studies, carried out the experiments and wrote the manuscript. G.R. discussed results and revised the manuscript.

COMPETING FINANCIAL INTERESTS

The authors declare no competing financial interests.

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METHODS

Strains. N2 Bristol was used as the wild-type strain. The following mutant strains were used: *rrf-3(pk1426)*, *daf-2(e1368)*, *daf-16(mu86)*, *eat-2(ad465)*, *skn-1(zu67)*, *glp-1(e2141)*, *lipl-1(tm1954)*, *lipl-3(tm4498)*, *mxl-3(ok1947)*, *mxl-3(tm2580)* and *hlh-30(tm1978)*. For *lipl* transcriptional fusions, *Is*[*lipl-1p*::GFP-pest], *Ex*[*lipl-2p*::RFP-pest;*myo-2p*::GFP], *Ex*[*lipl-3p*::RFP-pest;*myo-2p*::GFP] and *Ex*[*lipl-4p*::GFP;*myo-2p*::RFP] were used. For LIPL overexpression, *Ex*[*ges-1p*::LIPL-1cDNA::SL2::GFP;*myo-2p*::RFP], *Ex*[*ges-1p*::LIPL-2 cDNA::SL2::GFP; *myo-2p*::RFP] and *Ex*[*ges-1p*::LIPL-3 cDNA::SL2::GFP; *myo-2p*::RFP] were used. *ges-1* P driving *lipl-2* and *lipl-3* complementary DNAs were fused to TagRFP, and LIPL-1 to eGFP for subcellular localization experiments. LIPL-1::TagRFP is toxic, and LIPL-1::mGFP shows very dim signal. For tissue distribution, rescue analysis and ChIP of MXL-3, *mxl-3p*::MXL-3::mGFP;*myo-2p*::RFP was used. The *vha-6* promoter was used to drive intestinal tissue-specific rescue of *mxl-3*. For *mxl-3* tissue-specific knockdown analyses, lines were created following the strategy of ref. 29. Briefly, double stranded RNAi against *mxl-3* was specifically generated in the gut or AWC neurons of the non-spreading *sid-1(qt9)* strain expressing the *lipl-1* P::GFP transcriptional reporter. *Is*[*lipl-1p*::GFP-pest] was crossed into *sid-1(qt9)*. *Ex*[*ges-1p*::Mxl-3IR::SL2::GFP;*myo-2p*::RFP], *Ex*[*vha-6p*::Mxl-3IR::SL2::GFP;*myo-2p*::RFP], *Ex*[*odr-1p*::Mxl-3IR::SL2::GFP;*myo-2p*::RFP], *Ex*[*odr-3p*::Mxl-3IR::SL2::GFP;*myo-2p*::RFP] and *Ex*[*mxl-3p*::Mxl-3IR::SL2::GFP;*myo-2p*::RFP] were created by cloning the desired promoters in the AscI/Pme sites of pWormgatePro plasmids and then using the LR/BP recombinase Gateway system to introduce inverted repeat sequences of the *mxl-3* open reading frame. The constructs were injected into *sid-1(qt9)* animals carrying *lipl-1* P::GFP. For tissue distribution, rescue and ChIP analysis of HLH-30, *hlh-30p*::HLH-30cDNA::eGFP::*hlh-30* 3'UTR; *myo-2p*::RFP was used. DA2123 was used to assess autophagy levels; the transgene was crossed into *mxl-3(ok1947)* and *hlh-30(tm1978)*. Strain VS20 was used in ATGL-1 localization analyses.

***C. elegans* fasting assay.** *Transcriptional analyses.* Young adults (adult vulva with fewer than five eggs) were harvested, washed in a 35 µm nylon mesh, and seeded in empty NGM plates (fasting) or back onto OP50 plates (well-fed control). After the fasting period, treated and controls were harvested, washed in a 35 µm nylon mesh, and quickly frozen in liquid nitrogen. qRT–PCR data presented are from at least three independent experiments and all values are normalized to *ama-1* as internal control as well as to transcript levels in the untreated or wild-type animals.

Fat content assessment. Except for *hlh-30(tm1978)* experiments, hatchlings were synchronized for 48 h on S-basal, then seeded on OP50 plates, and incubated at 20 °C, unless otherwise stated. Animals were fasted for the indicated amounts of time and immediately treated for ORO staining[13], fixed for electron microscopy, or frozen for later processing. FAMEs were extracted and measured as previously reported[30]. Equal numbers of worms were compared.

Starvation survival. Approximately 10,000 synchronized hatchlings obtained by egg preparation and incubation in minimal medium, or synchronized young L4s extensively washed through a 20 µm mesh, were followed for each strain. No difference in survival was observed between *hlh-30(tm1978)* and wild-type worms if eggs were seeded immediately after bleaching on food-containing plates, suggesting that *hlh-30(tm1978)* mutant worms are not hypersensitive to bleach. Hatchlings or larvae were resuspended in 10 ml S-basal to which 2.5 µl 5 mM Sytox Green (Molecular Probes) and 1 µl 10% Triton X-100 were added. The worm suspensions were maintained rocking at 20 °C. A minimum of 500 worms were run through a COPAS Biosort (Union Biometrica) at the indicated times. Animals with green fluorescent signal of 50 or over were dead, as previously established by lack of movement in a liquid drop. Percentage survival $= (1 - (\text{worms with green signal} \geq 50/\text{total worms})) \times 100$.

Mammalian fasting assays. *Mouse experiments.* Five 9-week-old females were fasted for 10 h (8 pm–6 am), and five were fed *ad libitum*. Liver samples were extracted and immediately frozen in liquid nitrogen. RNA was extracted according to manufacturer's recommendations (TriReagent, MBP). Cyp4a14 was used as positive control and actin B to normalize cDNA. Each experiment was repeated twice. Lipase activity from whole lysates was measured in a reaction mix as described below.

HepG2 experiments. HepG2 human hepatocytes were grown in IMDM complete medium supplemented with 10% heat-inactivated fetal calf serum. At 50% confluence, three plates were washed with PBS and the cells were incubated in serum-free Earle's balanced salt solution, whereas another three plates were left in complete media as controls. At the indicated times cells from one complete medium and one salt solution plate were harvested and frozen in liquid nitrogen. RNA was extracted according to manufacturer 's recommendations (TriReagent, MBP). Specific TaqMan probes were purchased from Applied Biosystems. insulin-like growth factor-binding protein (IGFBP) was used as a positive control, and actin B to normalize cDNA. Each experiment was repeated three times. TFEB was knocked down in HepG2 cells using human TFEB Stealth RNAi siRNAs (HSS111870, Invitrogen). Twenty-four hours after transfection using Lipofectamine RNAiMAX transfection reagent (Invitrogen), two plates of TFEB knocked-down cells and two plates of negative-control transfected cells (Stealth RNAi siRNA Negative Control) were harvested and split into a total of seven plates. Forty-eight hours after transfection, half the TFEB knocked-down and negative control cells were resuspended in ~~EEBS~~. At the indicated times, cells from one TFEB knocked-down complete medium and one TFEB knocked-down ~~EEBS~~ plus one negative control complete medium and one negative control ~~EEBS~~ plate, were harvested and processed for transcriptional analysis as described above. TFEB expression was tested by TaqMan PCR.

Lipase assay. Animals were resuspended in 300 µl of 200 mM sodium acetate buffer pH 4.5, and immediately frozen in liquid nitrogen. To assay, samples were sonicated and protein concentrations were measured. Reaction mix: 250 µg of total protein (up to 100 µl of lysate), 20 µl 2 M sodium acetate buffer at pH 4.5, 10 µl 4-methylumbelliferyl palmitate (4 mg ml^{-1}), bring reaction volume to 200 µl with water. Fluorescence was read after 2 h at room temperature in a plate reader, excitation 355 nm–emission 460 nm.

Immunostaining. GFP was revealed with anti-GFP antibody (Roche, catalogue no 11814460001) and PGP-2 was revealed with anti-PGP-2 antibody kindly provided by G. Hermann[11]. TagRFP antibody was raised in house.

RNAi screen. An RNAi sublibrary containing 403 predicted transcriptional regulators and co-regulators and 193 nuclear hormone receptors (Supplementary Table S2) was built by cherry-picking clones from the Ahringer genome-wide and the Vidal RNAi library. The strain carrying the GFP transcriptional fusion to *lipl-1* was used to screen for transcriptional regulators that affect transcriptional activation of lipolysis. RNAi bacteria were cultured for 12 h in Luria–Bertani medium with 100 µg ml^{-1} ampicillin, and then washed with S-basal and seeded onto RNAi agar plates containing 5 mM isopropyl-β-D-thiogalactoside (IPTG). The plates were left to dry in a laminar-flow hood and incubated at room temperature overnight to induce dsRNA expression. Synchronized *Is*[*lipl-1p*::GFP-pest; *myo-2p*::RFP] hatchlings were seeded on the plates. After 3 days of incubation at 20 °C, young adults were scored for GFP signal (animals fed empty vector control showed no signal or a dim signal limited to the tail in these conditions).

Yeast one-hybrid analyses. The ability of MXL-3 to bind to the promoters of the lipases was tested in the yeast one-hybrid system (Y1H), according to Deplancke *et al.* with some modifications. Briefly, *lipl-1* to *4* promoters (3 kb upstream up to the ATG) were fused to β-galactosidase or HIS3 and integrated in YM4271. Two clones of each promoter that did not grow in 25 mM 3-aminotriazole (3AT) and showed a pale blue colour in the β-gal assay were transformed with only the Gal-4 activation domain or the activation domain fused to the MXL-3 open reading frame. Twenty-four random colonies from each transformation were picked and tested for survival in 3AT or β-gal signal. Expression of a transcriptional regulator able to bind to the tested promoters renders yeast resistant to 3AT and β-gal positive.

ChIP-qPCR. qPCR of DNA immunoprecipitated from well-fed or 6 h-fasted transgenic animals expressing MXL-3::GFP or HLH-30::GFP was carried out as previously described[31]. The MXL-3::GFP line used expresses 4.2 ± 0.38 times more *mxl-3* than wild-type worms. The HLH-30::GFP line expresses 6.75 ± 0.91 times more *hlh-30* than wild-type worms. Monoclonal mouse αGFP antibody from Roche (catalogue no 11814460001) was used for immunoprecipitation . Three sets of primers surrounding the CACGTG target sites up to 500 bp from the start site of *lipl-1* or *lipl-3* genes were used in qPCRs to compare the immunoprecipitated to input DNA. Fold change was calculated relative to the amplification obtained using two sets of primers surrounding the CACTAT sequence at −88 bp of the *ama-1* start site from immunoprecipitated and input DNA.

Electron microscopy. Transmitted electron microscopy of osmium-, propylene oxide- and EPON-treated wild-type or mutant young adults was carried out in the Microscopy Core of the Center for Systems Biology/Program in Membrane Biology, which is partially supported by Inflammatory Bowel Disease Grant DK43351 and Boston Area Diabetes and Endocrinology Research Center (BADERC) Award DK57521.

LRO content assessment. Animals were synchronized through a 2 h synchronous egg lay or by 24 h hatching at 20 °C in S-basal minimal media. Worms were seeded on NGM plates containing *E. coli* OP50 supplemented with 1 µM of the fluorescent

fatty acid analogue C1-BODIPY 500/510-C12 (Invitrogen) or 25 ng ml^{-1} of Nile red. Imaging and quantification was conducted using an Axioplan microscope and Axiovision software (Zeiss). At least 25 animals were imaged in at least two independent experiments.

Lysosomal fractionation. Lysosomes were isolated from mixed stage LIPL-3::TagRFP animals using a LYSISO1 kit (Sigma) as previously described[32]. The lysosome enrichment was determined by measuring acid phosphatase activity (BioVision) normalized to protein concentration. LIPL-3::TagRFP was detected using α-TagRFP antibodies from Evrogen. Acid lipase activity was measured as described above.

Longevity assays. Synchronized day-1 adults were transferred to fresh NGM OP50 or RNAi plates every two days until no progeny was observed and then once a week for RNAi to keep the strength of the treatment. Two to six independent assays were carried out. Lifespan experiments were conducted at 20 °C, unless otherwise stated. Kaplan–Meier survival analysis was done using SSPS 17 software; significance was determined using log-rank statistics.

Statistical analyses. *t*-test analyses were carried out in all experiments with three or more biological replicates (independent experiments). For experiments with $n = 2$, representative data are presented. Log-rank statistics was used for the analysis of the longevity experiments as explained above.

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Query 1:
Please check the email address given for corresponding author.

Query 2:
Please provide post code for both affiliations and also provide more details for 1st affiliation.

Page 2

Query 3:
Please provide text to define ddCts.

Page 4

Query 4:
'TOR' expanded as 'target of rapamycin'—please check, and correct if necessary.

Page 7

Query 5:
Please provide text to define AWC.

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Query 6:
Please confirm statement: 'The authors declare no competing financial interests.'

Query 7:
Please provide year for ref. 16.

Query 8:
References 30 and 32 have been deleted from reference list since they were the repetition of refs 13 and 11 respectively and hence subsequent references renumbered and citations also changed. Please check.

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Query 9:
'IGFB' changed to 'insulin-like growth factor-binding protein (IGFBP)'—please check, and correct if necessary.

Query 10:
'LB' expanded as 'Luria–Bertani medium'—please check, and correct if necessary.

Query 11:
Is 'Deplancke et al' ref. 31?

General Queries

Query 12:
For the representation of gene symbols and genotypes we follow the standard scientific conventions and nomenclature found in databases such as HUGO for humans, MGI for mice or Flybase for Drosophila. Accordingly, many changes may have been made throughout the text and figures. Please check that we have interpreted each instance correctly.

Query 13:
Please note that reference numbers are formatted according to style in the text, so that any reference numbers following a symbol or acronym are given as 'ref. XX' on the line, whereas all other reference numbers are given as superscripts.

Query 14:
'UTR' expanded as 'untranslated region' in caption of Fig. 3.—please check, and correct if necessary.

Query 15:
Please give 'LRO' in full in caption of Fig. 5.

Query 16:
Please provide text to define EEBS in caption of Fig. 6.